



Institut luxembourgeois de la normalisation  
de l'accréditation, de la sécurité et qualité  
des produits et services

**ILNAS-EN 17805:2023**

**Water quality - Sampling, capture and  
preservation of environmental DNA  
from water**

Qualité de l'eau - Échantillonnage,  
collecte et conservation de l'ADN  
environnemental prélevé dans l'eau

Wasserbeschaffenheit - Probenahme,  
Erfassung und Konservierung von  
Umwelt DNA in Wasser

**03/2023**



## National Foreword

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## Water quality - Sampling, capture and preservation of environmental DNA from water

Qualité de l'eau - Échantillonnage, collecte et conservation de l'ADN environnemental prélevé dans l'eau

Wasserbeschaffenheit - Probenahme, Erfassung und Konservierung von Umwelt-DNA in Wasser

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EUROPÄISCHES KOMITEE FÜR NORMUNG

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## Contents

Page

European foreword.....	3
Introduction .....	4
1 Scope .....	5
2 Normative references .....	5
3 Terms and definitions .....	5
4 Principle .....	7
5 Procedure.....	8
5.1 General.....	8
5.2 Considerations prior to fieldwork .....	8
5.3 Equipment preparation prior to fieldwork .....	8
5.4 Sampling the eDNA from water.....	8
5.5 Preserving the sample.....	9
6 Equipment .....	10
7 Preservative solutions.....	11
7.1 General.....	11
7.2 Examples of preservative solutions .....	11
8 Sampling report.....	12
8.1 General.....	12
8.2 Sample identity and characteristics.....	12
8.3 Sampling site.....	12
8.4 Sampling conditions.....	12
8.5 Sampling.....	13
9 Avoiding sample contamination .....	13
9.1 General.....	13
9.2 Contamination that originates from equipment.....	13
9.3 Sampling equipment decontamination procedure.....	14
Annex A (informative) Filter types .....	15
Bibliography.....	16

## European foreword

This document (EN 17805:2023) has been prepared by Technical Committee CEN/TC 230 “Water analysis”, the secretariat of which is held by DIN.

This European Standard shall be given the status of a national standard, either by publication of an identical text or by endorsement, at the latest by September 2023, and conflicting national standards shall be withdrawn at the latest by September 2023.

Attention is drawn to the possibility that some of the elements of this document may be the subject of patent rights. CEN shall not be held responsible for identifying any or all such patent rights.

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## Introduction

**WARNING — Persons using this document should be familiar with water sampling protocols to assess biological diversity. This document does not purport to address all of the safety problems, if any, associated with its use. It is the responsibility of the user to establish appropriate health and safety practices.**

**Moreover, the need of notification, obtaining certificates or permits prior to sampling, depending on national or international laws and regulations such as the Nagoya Protocol on Access to Genetic Resources (<https://www.cbd.int/abs/>), needs to be considered.**

The monitoring of organisms is key to the assessment of the status of aquatic ecosystems and is required by national and international legislation such as the European Union Water Framework Directive (2000/60/EC). A range of methods have been described how to monitor organisms in aquatic environments, leading to a wide range of European standards (e.g. EN 14011:2003, EN 14757:2015, EN 15460:2007). These approaches, however, necessitate the capture and/or collection of the organisms of interest, which can be a laborious and time-consuming process.

The possibility to detect the presence of organisms and/or quantify relative abundance (e.g. [6]) in aquatic environments via the analysis of environmental DNA (eDNA) provides a novel means to monitor biodiversity across a wide range of taxonomic groups, including microorganisms, plants and animals ([7][8][9]). This approach allows to examine organismic diversity without the need to directly isolate and capture organisms and it is expected to play a key role for future biomonitoring aiming at temporally and spatially highly resolved species inventories [10]. Albeit the power of the eDNA approach has been repeatedly reported [11], there is a great need for standardizing the application of eDNA-based assessment of aquatic biodiversity ([12], [13]). Note, however, that eDNA-based biomonitoring currently does not allow to obtain certain population parameters (e.g. individual size, sex) which can be obtained by traditional sampling techniques.

This document provides guidance how to sample and preserve eDNA from water samples, addressing the first and crucial step for any further downstream eDNA-based analyses of biodiversity. A specific technical report for the routine sampling of benthic diatoms from rivers and lakes adapted for metabarcoding analyses is CEN/TR 17245:2018.

## 1 Scope

This document specifies procedures for sampling, capture and preservation of environmental DNA (eDNA) in aquatic environments, stemming from organisms that are or have recently been present in a waterbody, have visited it or whose DNA has been introduced to the waterbody through some mechanism. This document also covers procedures for avoiding sample contamination and ensuring DNA quality, key properties of the filtering procedure and equipment and reporting standards.

This document does not include the collection of eDNA from biofilms, sediments or similar sample types and does not cover sampling designs.

## 2 Normative references

There are no normative references in this document.

## 3 Terms and definitions

For the purposes of this document, the following terms and definitions apply.

ISO and IEC maintain terminology databases for use in standardization at the following addresses:

- IEC Electropedia: available at <https://www.electropedia.org/>
- ISO Online browsing platform: available at <https://www.iso.org/obp>

### 3.1

#### **cross-contamination**

unintended transfer of any source of and/or DNA from one sample to another sample

### 3.2

#### **decontamination**

procedure to remove any source and/or trace of DNA from material that might come into contact with the sample

### 3.3

#### **enclosed filter**

filtering device where the filter membrane is encapsulated and where the inflow and outflow can be closed for transport and storage

Note 1 to entry: The eDNA contained on the filter is typically extracted without removing the membrane from the filter capsule greatly reducing the risk of contamination of samples. See Figure A.1 C. in Annex A.

### 3.4

#### **environmental DNA**

##### **eDNA**

material stemming e.g. from dead or from living organisms and include single-stranded (ss) and double-stranded (ds) DNA fragments from nuclear and mitochondrial/plastid DNA of eukaryotes as well as plasmid DNA of prokaryotes

Note 1 to entry: Subsuming DNA from various sources such as unicellular or small multicellular organisms or tissue particles (e.g. shed cells, faeces) and gametes of multicellular organisms.

### 3.5

#### **field equipment blank**

sample obtained from processing target DNA-free water (e.g. distilled water) through all the equipment used and covering all procedures involved in the eDNA sampling process to allow checking that the equipment and procedures do not introduce DNA contamination

### 3.6

#### **housed filter**

systems in which a filter membrane is protected within a solid housing during the filtration process

Note 1 to entry: The filters are removed from the housing for eDNA extraction. The housing can be opened and the filter removed for preservation and later processing. See Figure A.1 B. in Annex A.

### 3.7

#### **lysis buffer**

buffer solution to preserve DNA present in the sample and to lyse/open cells as a first step of the DNA extraction

### 3.8

#### **internal positive control**

##### **IPC**

known fragment of synthetic or natural DNA containing an amplifiable and quantifiable sequence that will not naturally occur in the sample

Note 1 to entry: The IPC can be added to the sample or the preservation/lysis buffer at a known concentration to verify the efficiency of DNA preservation, DNA extraction, DNA amplification and DNA identification.

### 3.9

#### **open filter**

filtering device including filtration towers (laboratory) and filtration backpacks from which the filter membrane has to be removed by hand for further processing

Note 1 to entry: See Figure A.1 A. in Annex A.

### 3.10

#### **pre-filter**

filter membrane, mesh or hose strainer with a larger pore-size than the main filter membrane (for capturing the eDNA) through which water is passed first to remove larger particles of sediment, plant material or algae to increase the volume of water that can be filtered before saturation of the main filter

### 3.11

#### **sample contamination**

process by which exogenous DNA is unintentionally introduced to the sample during the sampling process

Note 1 to entry: DNA that is already present in the water before the eDNA sampling was undertaken is not considered as contamination.

### 3.12

#### **target DNA**

any source and/or trace of DNA from the surveyed species/taxa